**IDEXX Quality Starts with Quality Samples**

The quality and accuracy of results are very important to everyone at IDEXX

**IDEXX Tips for obtaining high quality samples:**

**Choose the correct sample and container**
Read the collection guidelines on the following pages or in the Sample Requirements column by each test in the Reference laboratory section of the current edition of the Directory of Products and Services (call 00800 1234 33 99 if you cannot find the test you require) before taking a sample. Improper choice of a collection vial can adversely affect results.

**Did You Know?** Using a serum gel tube is an easy way to collect a good serum sample for chemistry panels. However, for certain specialised tests, such as many endocrinology and drug tests, the gel can interfere with results.

**Did You Know?** Whole blood EDTA samples start to degrade as soon as the blood is outside of the animal.

To preserve cell morphology, include air-dried, unstained blood slides along with the EDTA tube. Slides should also be included with samples submitted for fluid analysis for accurate cytologic interpretation. Blood smears made at the time of blood collection help avoid platelet clumping problems and allow more accurate platelet interpretation.

**Did You Know?** Including slides with your Comprehensive Haematology can help with pathologist reviews of unusual cells, and with the identification of red blood cell parasites.

**Use proper techniques**
Filling syringes: When filling syringes, aim for a good free flowing stick from the largest accessible vein. Slow draws and difficult sticks can rupture red cells, adversely affecting CBC results and certain chemistries. The longer the blood stays in the syringe, the greater the risk of clumped platelets and clots that degrade test results. Always fill the tubes without additives first. This prevents carry-over of tube additives. For example, if you are filling an EDTA tube and a serum gel tube, always fill the gel tube first. Even a small amount of EDTA can interfere with many chemistry results. Fill EDTA or citrate tubes precisely. Overfilling and under-filling tubes causes the wrong ratio of additives. Excess EDTA in an under-filled tube will give inaccurate CBC results. Overfilling the EDTA tube may cause the sample to clot. Centrifuging: Make sure samples are fully clotted before tubes are centrifuged.

Note that some samples may take longer to clot than others.

**Did You Know?** In tests that require serum, it is important to collect and centrifuge the sample properly. If a tube is spun too soon after drawing the blood, you will send plasma to the laboratory and not serum.

If you have any questions regarding submission of specimens, or if you require appropriate supplies, please call us on 00800 1234 33 99.
Specimen Collection Guidelines

**Label all tubes and slides submitted with:** Animals Name, Owners Name, Date of Sampling and Time of Sampling

<table>
<thead>
<tr>
<th>Type of Testing</th>
<th>Specimen</th>
<th>Container</th>
<th>Contents</th>
<th>Protocol</th>
<th>Storage</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Chemistry, Immunology, Endocrinology</strong></td>
<td>Serum</td>
<td>Serum Gel Tube</td>
<td>Get to separate serum from clot (during centrifugation) and a clot activator</td>
<td>Let specimen clot 15 - 20 minutes, centrifuge at 2,500 rpm for 10 - 15 minutes. DO NOT use serum gel tubes for therapeutic drug monitoring (digoxin, phenobarbital), or Teppoton HS I</td>
<td>Refrigerate</td>
</tr>
<tr>
<td><strong>Chemistry (including Teppoton HS I, Immunology, Endocrinology including progesterone and therapeutic drug monitoring)</strong></td>
<td>Serum</td>
<td>Serum Tube</td>
<td>No additives (simply/sterile)</td>
<td>Let specimen clot 15 - 20 minutes, centrifuge at 3,000 rpm for 10 - 15 minutes, remove serum and transfer to a plain serum tube</td>
<td>Refrigerate</td>
</tr>
<tr>
<td><strong>Chemistry Cardiopet® proBNP</strong></td>
<td>0.5 ml separated EDTA plasma</td>
<td>Plain tube</td>
<td>No additives (simply/sterile)</td>
<td>Collect specimen in an EDTA tube, Centrifuge sample tubes (10 minutes at 3000 rpm or StartSpin VT hard cycle). Use a pipette to transfer plasma to plain sample tube labelled BIPE Carcin/Feline. NOTE: The Cardiopet® proBNP assay is stable for 48 hours at room temperature. Please submit samples by courier or Guaranteed Mail. If you anticipate a delay in transportation, please submit samples cooled on ice.</td>
<td>Keep frozen</td>
</tr>
<tr>
<td><strong>Haematology</strong></td>
<td>Whole blood</td>
<td>EDTA Tube</td>
<td>Anticoagulant EDTA</td>
<td>Filling tube to the line mark on the tube will allow to obtain the correct blood to anticoagulant ratio. Invert gently several times after filling</td>
<td>DO NOT freeze</td>
</tr>
<tr>
<td><strong>Coagulation (PT, APTT, fibrinogen, anti-thrombin III, D Dimer, (PT, APTT, fibrinogen, Factor VII, Phenobarbital))</strong></td>
<td>Citrated Plasma</td>
<td>Citrate Tube</td>
<td>Anticoagulant Sodium citrate</td>
<td>Correct blood to anticoagulant ratio is very important. Take blood sample from animal. At least 2 ml of blood is required to run PT and APTT (yield minimum of 1 ml citrated plasma). Remove needle and fill sample tubes in the appropriate order and to the level indicated on each tube. Mix anti-coagulated tubes. Ensure the coagulation tubes are not clotted. If they are, repeat sampling is required. Centrifuge coagulation sample tubes (10 minutes at 5000 rpm or StartSpin VT hard cycle). Use a pipette to transfer plasma to plain sample tube labelled CIPTT Plasma. Please remember to include an EDTA sample for platelet count, which is part of our coagulation profile. To ensure the best results, please submit the citrated plasma samples frozen, packed with ice packs to prevent freezing, by overnight delivery to the laboratory. NOTE: Samples that are clotted or haemolysed will not be analyzed because they would fail to provide accurate results.</td>
<td>Keep frozen</td>
</tr>
<tr>
<td><strong>Avian Blood and Miscellaneous chemicals</strong></td>
<td>See Avian and exotic specimen submission guidelines in this section. Check individual test listing.</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Urine Analysis</strong></td>
<td>Urine</td>
<td>Urine collection preferred</td>
<td>Plain urine container</td>
<td>After collection, cap container.</td>
<td>Refrigerate and prevent UV/Sunlight exposure</td>
</tr>
<tr>
<td><strong>Microbiology</strong></td>
<td>See Microbiology Specimen Submission Guidelines</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Cytology</strong></td>
<td>See Cytology Specimen Submission Guidelines</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Biopsy</strong></td>
<td>See HistoPathology Specimen Submission Guidelines</td>
<td></td>
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<td></td>
</tr>
</tbody>
</table>

**Normal Flora, Predictable Susceptibility Patterns and Non-pathogenic Organisms**

*CHECK* follows guidelines set by CSLI, combined with our years of experience in performing susceptibility testing. We believe these are “best practice” microbiology techniques, and would be happy to discuss the following policies with you.

**Sterile Tubes**

Use glass or plastic tubes with no additives. Serum Gel tubes with clot activator are not acceptable for cultures because the clot activator binds bacteria, which inhibits growth.

**Fluids**

Make sure all collection devices containing fluids are sealed and leak proof before submitting. NOTE: Specimens that are >48 hours old are not suitable for culture, and loss of viability should be expected.

**Blood Culture**

Aerobic and anaerobic cultures are performed on all blood cultures.

<table>
<thead>
<tr>
<th>Type of Source</th>
<th>Collection Device</th>
<th>Specimen Preparation and Collection</th>
<th>Test to Request</th>
</tr>
</thead>
<tbody>
<tr>
<td>Abscess or Wound</td>
<td>Transport Swab</td>
<td>Aseptically prepare collection site. Aspirate fluid or pus from pustules or vesicular wounds and abscesses.</td>
<td>Aerobic and Anaerobic Culture</td>
</tr>
<tr>
<td>Blood</td>
<td>2 – 5 ml for small animal 10 ml for large animal</td>
<td>Wash puncture site with antiseptic soap; sterilize for platelet count, which is part of our coagulation profile. To ensure the best results, please submit the citrated plasma samples frozen, packed with ice packs to prevent freezing, by overnight delivery to the laboratory. NOTE: Samples that are clotted or haemolysed will not be analyzed because they would fail to provide accurate results.</td>
<td>Blood Culture</td>
</tr>
<tr>
<td>Bone Marrow</td>
<td>Plain sterile container</td>
<td>Aseptically prepare collection site.</td>
<td>Aerobic and Anaerobic Culture</td>
</tr>
<tr>
<td>Central Nervous System</td>
<td>Plain sterile container</td>
<td>Collect CSF fluid by an aseptic subdural tap, or wound aspiration or lumber puncture.</td>
<td>Aerobic and Anaerobic Culture</td>
</tr>
<tr>
<td>Ears</td>
<td>Transport Swab</td>
<td>Use swab to collect suppurative material from cul-de-sac or middle ear.</td>
<td>Aerobic Culture</td>
</tr>
<tr>
<td>Eyes</td>
<td>Transport Swab</td>
<td>Use swab to collect suppurative material from cul-de-sac or medial canthus.</td>
<td></td>
</tr>
</tbody>
</table>
Cytology

Specimen Collection Guidelines

**ACCURATE RESULTS DEPEND ON QUALITY SPECIMENS. PLEASE FOLLOW THESE GUIDELINES:**

Perform sampling by fine-needle aspiration or non-aspiration biopsy, scrapings or imprints. Prepare slides in-clinic using either a “squash” preparation or blood-smear technique. Call if you have questions about slide preparation. Please DO NOT submit syringes with needles.

Patient history and clinical findings contribute to an accurate result.

**On containers and slides, please write:**
- Patient’s name
- Site/Source

**On the requisition form, include:**
- Patient details (owner’s name, patient’s name, age, sex, species, breed, etc.)
- Reference to any previous laboratory results (FBC, biochemistry profile, prior cytology/histology or serology) be sure to include our laboratory reference numbers
- Gross lesion description
- Specific anatomic location (e.g., cutaneous, subcutaneous, deep tissue, intra-thoracic, intra-abdominal)
- Size, shape, consistency, symmetry, definition of borders
- Clinical history – duration of lesion, progression of lesion, treatment and response to therapy
- Radiographic and ultrasonographic findings

**When submitting aspirates and impressions:**
- Submit one to three unstained air-dried slides
- Store at room temperature
- DO NOT spray with hairspray or other fixatives
- DO NOT expose to formalin fumes
- DO NOT ship slides for cytology in the same bag as a formalin-containing tissue samples

**When submitting fluids and washes:**
- Enclose fluid in a plain EDTA tube along with unstained air-dried slides
- Prepare slides immediately after fluid collection to preserve cell morphology (most fluids are stable for only a few hours at room temperature).
- If volume allows, also supply fixed fluid in an EDTA Tube by adding two drops of 10% formalin saline (as supplied in our histology pot) per ml of collected liquid. Formalin fixation may improve cell preservation in fluids and is particularly indicated for the evaluation of cerebro-spinal fluid, respiratory specimens, and urine.
- DO NOT submit fluids in a gel tube, in a syringe, or as cover-slipped and wet preparations. Submission of fluid in a gel tube can interfere with accurate cytologic evaluation due to the presence of clotting activators.

**When submitting urine samples:**
- Please do not submit sedistain urine slides as these dry out and are non-diagnostic

**Histopathology**

**Specimen Collection Guidelines**

**ACCURATE RESULTS DEPEND ON QUALITY SPECIMENS. PLEASE FOLLOW THESE GUIDELINES:**

**Turnaround Time**
Most evaluations will be completed within 3 – 4 days of receipt in our laboratory (unless otherwise indicated). Additional fixation or decalcification will take longer. We will notify you if an unusually long delay is anticipated.

**Collection Technique**
Samples are collected for histological examination by standard surgical techniques or at post mortem examination.

**Labelling Criteria**
Please ensure all specimen containers are labelled with
- Patients name, date
- Type of specimen, (Site / Source)

**Requisition Information**
A thorough clinical history and details of the physical examination are essential for the correct histological interpretation of tissue changes. Information required includes details (species, breed, age, sex), a description of the appearance and distribution of lesions, duration of the condition, biopsy sites or post mortem tissue, response to prior treatments, current treatment regimes and any other relevant information. You may include any questions to be answered on your requisition form.

**Fixation Guidelines**
Tissue samples should be fixed in 10% buffered formalin.

**Did You Know?**
The 10:10:10 Rule? For optimum fixation and sectioning use 10% formalin; 10:1 ratio, and a biopsy size 10mm cubed.

- Place specimens in a biopsy pot supplied by IDEXX Laboratories, with the ratio of formalin to tissue > 10:1.
- Samples that are very tiny (less than 2mm in diameter) and friable, haemorrhagic, mucoid or fatty in nature may not be suitable for histology processing and interpretation. If a biopsy of this size or nature is received the lab will proceed with testing but inform you if the sample submitted is potentially inadequate. If the sample is inadequate you will be informed via a preliminary report, that the tissue did not survive processing or is not diagnostically viable, upon which the charge will be removed.
- With the exception of biopsies, if submitting multiple samples please use separate pots for each specimen and label each with anatomical site. If samples are to be placed together, please clearly indicate this on the request form, and refrain from submitting small samples (under 1cm) being submitted in the same container as larger ones.
- Submit entire lesions and tumours with adjacent excised tissue.
- For rapid fixation of larger lesions and tumours, cut a section 0.5-1cm wide through the centre of the specimen. Where possible, cut the skin surface, so the deep surgical margin is not cut.
- Open hollow organs, such as intestine, prior to placing them in fixative.
- Small fragile specimens (bone marrow, Tu-cut liver or kidney) can be submitted in mesh cassettes, available from our supplies department.

**Transport Guidelines**
All samples should be placed in a well sealed leak proof bag containing enough absorbent material for the volume of formalin. Fixed tissue which is to be mailed may be placed in a leak proof plastic bag or container with a formalin soaked gauze to keep the tissue moist (ensure adequate fixation has occurred prior to transportation).

**Necropsy Samples**
IDEXX no longer offers in-laboratory necropsy service, but there are a variety of options for submitting necropsy samples. Please contact our Customer Support on 00800 1234 3399 for further advice.
Accurate results depend on quality specimens

Blood samples should be collected aseptically from veins, most commonly the jugular vein in companion birds. Ulnar veins or tibiotarsal veins can be used in some species. Nail clipping is NOT a suitable method for collecting blood samples.

**Biochemistry Only**
Submit sample in lithium heparin tubes. Gel separator tubes can be used. Gel tubes can be spun before submission, and this will reduce storage artefacts. Plain serum tubes can also be used, but result in a smaller sample for analysis, and may reduce the number of measurements that can be performed.

Small sample volume may limit the number of biochemistry tests that can be performed. If submitting a small sample, please note your required tests in order of preference on the submission form.

**Haematology Only**
Submit slide(s) and whole blood. Make slides immediately after collection, using blood that has not been exposed to anticoagulant. Preferably make two slides, one by the usual slide-and-slide method, one by a coverslip-and-slide method. Air dry slides immediately, but do not fix or stain. Submit the whole blood in lithium heparin. Gel separator tubes can be used but must NOT be spun down. EDTA can also be used for most birds (excluding ratites, crows and ravens), but is not the preferred sample. EDTA should not be used for reptile blood, use of lithium heparin is recommended.

**Haematology and Biochemistry Required**
Submit blood films (see haematology above) and blood in lithium heparin. Do NOT spin gel separator lithium heparin tubes if only a single tube is submitted. At least one unspun lithium heparin tube must be submitted. Small sample sizes may limit the number of biochemistry tests that can be performed. If submitting a small sample, please note your required tests in order of preference on the submission form.

**Spoiled Samples**
When a sample is received in poor condition, e.g. clotted, lipaemic, or haemolysed, we will perform the requested tests wherever possible. Many spoiled samples can be avoided by the use of correct tubes. It is for this reason that we provide polypropylene, screw top tubes, free of charge. (Shipping charge of £7.50 + VAT will apply to each dispatch)

**Samples from patients on chemotherapy treatment**
Please indicate clearly on the submission form if samples are from patients on chemotherapy treatment. Cytotoxic drugs are eliminated by renal or faecal excretion within 48 hours of being administered, and all samples require special handling in order to comply with health and safety regulations.

**Sample Tubes & Supplies**
Sample tubes are provided free-of-charge by IDEXX Laboratories. These tubes are designed to be used with our analysers, and use of other tubes may cause delay in processing.

- **EDTA Tubes**
- **Serum Tubes**
- **Citrate Tubes**
- **Heparin Tubes**
- **Swabs with charcoal media**
- **Plain Swabs (for PCR tests)**
- **CEM Swab**
- **Plain Universal Containers**
- **Boric Acid Universal Containers**
- **Faecal Universal Containers**
- **Histology Pots containing 10% formalin**
- **Endoscopy Biopsy mesh Cassettes**
- **Slide Containers**
- **Packaging Supplies**

All of these tubes are supplied free of charge to our clients (shipping charge of £7.50 + VAT for each order). To order supplies, please fax one of our supplies order forms to 01937 544 170 or order online at idexx.co.uk

**Avian and Exotic**

The following guidelines will allow our staff to process your specimens safely, efficiently and accurately. They will also help to ensure your samples arrive with us in a satisfactory condition for testing.

**Please:**
- Do not use tubes or swabs that are past their expiry date.
- Do not send unlabelled tubes.
- Do not stick labels onto tubes – we need to bar code all specimens on receipt and this is hindered by this practice.
- Do not cover the tubes with sticky tape – if the lids are screwed on tightly they will not leak.
- Do not send hypodermic needles.
- Do not send glass tubes or slides unless suitably packaged. Slide holders are available free-of-charge from our laboratory.
- Do not send any sample unless suitably packaged – specific packaging instructions are provided and packaging material is available from our laboratory. It is the sender’s responsibility to ensure that all pathological specimens are packed in accordance with current Royal Mail or Courier regulations. Non-adherence to packaging instructions may result in sample return or disposal by the carrying company and the imposition of possible financial penalties.

**Please note:** We keep all specimens received at the laboratory for seven (7) days after which they will be disposed of in an appropriate manner, so when the results of your initial sample submission point to the next step in your investigation, and we have the appropriate sample at our laboratory, simply call our Customer Support Team to discuss your requirements.

**Nail clipping** is NOT a suitable method for collecting blood samples.

**Blood samples should be collected aseptically from veins, most commonly the jugular vein in companion birds. Ulnar veins or tibiotarsal veins can be used in some species. Nail clipping is NOT a suitable method for collecting blood samples.**

All enquiries:
- 08000 1234 33 99
- 01937 544 001
- labhelp@idexx.com
Regulations for the shipment of Pathological Samples to the Laboratory

- All known infectious samples must be packaged in accordance with IATA602 in approved IARA602 infectious boxes
- Diagnostic samples must be packaged in accordance with the revised Royal mail regulations

According to the IATA regulations, Infectious substances are defined as:

Infectious substances:
"Substances which are known to contain, or are reasonably expected to contain, pathogens. Pathogens are defined as micro-organisms (including bacteria, viruses, rickettsiae, parasites, fungi) and other agents such as prions, which can cause disease in humans or animals."

Classification of biological products and Diagnostic Specimens:
"Those known or thought likely to contain infectious substances. Diagnostic specimens being transported to undergo confirmatory testing must be considered to fall within this group (see packing instructions P650)."

You represent and warrant that any sample containing any hazardous substance which is to be delivered to IDEXX will be packaged, labelled, transported, and delivered in accordance with applicable laws.

Biological substances must be posted in packaging that complies with Packaging Instruction P650 and the senders name and return address must be clearly visible on the outer packaging. If you would like to view the comprehensive guidelines on how to package and submit samples to the Reference Laboratory, the IATA Guidance document is available on the Royal Mail website: http://www.royalmail.com/sites/default/files/Guidance-Document-Infectious-Substances-171012.pdf

If you wish to send samples that are known to be infectious, please ensure that the IATA 602 regulations are met.

Please note: we are unable to accept any sample falling into Category A – high risk to humans

Approved IATA 602 boxes can be purchased through a number of suppliers or can be purchased / rented at a discounted rate from IDEXX Laboratories.

The above information is a guide only and if you have any further queries please do not hesitate to contact our Customer Support team: 00800 1234 3399 Option 1 followed by Option 1

Packaging you Samples for Dispatch to the Reference Laboratory

STEP 1 – ORDER TEST KITS

1. Category B Shipping Box
2. Order Form
3. Specimen

STEP 2 – PREPARE BOX

4. Pressure Bag
5. IDEXX Overpack Bag

1. Assemble the Category B box by folding up the top half of the box. The side with the two cut out holes forms the bottom of the box.
2. Fold in the small side flaps so they are hovering over the holes.
3. Fold up the sides of the box and lock the two tabs into place in the notches in the base of the box.
Sample Dispatch Guidelines

**STEP 3 – PREPARE SAMPLES**

1. Ensure you complete the appropriate Requisition Form (remembering to include both your practice code and practice address) and submit all sample types required. Ensure that the sample tubes and slides are labelled with the patient and owners names, and your practice Vet Code.

2. Ensure the caps are tightly screwed onto each sample container, and wrap the tubes in a piece of absorbent material in case of leakage. Please do not use sellotape either on the tubes or on the packing material.

3. Place the wrapped tubes into the sealable pocket of the supplied Specimen Transport bag, and seal by removing the paper strip and holding the sticky tab over at the fold line.

4. Fold the Requisition Form with the typeface facing outwards (as in the picture) and place in the open pocket of the Specimen Transport bag.

**STEP 4 – SHIPPING STEPS**

1. Place the prepared Specimen bag(s) containing the sample(s) and order form(s) into the box.

   Please note: More than one specimen Biohazard bag can be placed in one shipping container, however only one requisition should be placed into the Biohazard bag.

2. Fold the sides of the lid into the box and push the top down.

3. Finally, fold in the side tabs and close the box.

4. Now that your box is prepared, place the box into the IDEXX Overpack bag and seal the bag by tearing off the red strip.

Following these guidelines will allow our staff to process your specimens safely, efficiently and accurately. They will also help to ensure your samples arrive with us in a satisfactory condition for testing.

If you would like any further advice please contact our Customer Support team:

Telephone: 00800 1234 33 99  Fax: 01937 544 001  e-mail: customersupportUK@idexx.com
Microbiology Request Form

Please ensure all text is legible. This helps us process your request more efficiently and also ensures you receive appropriate interpretation if you have requested an interpreted profile.

IDEXX Laboratories Ltd.

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Pathology services play a crucial role in ensuring accurate diagnosis and effective treatment and monitoring for patients. Our Wetherby Laboratory gained accreditation in 2004 by the United Kingdom Accreditation Service (UKAS) to BS EN ISO/IEC 17025:2000 (now updated to 17025:2006) General standards for the competence of testing and calibration laboratories and since then, with continuing process improvement, have expanded on the number of tests included in the scope of accreditation. The majority of tests in Biochemistry, Endocrinology, Haematology, Immunology, Microbiology, Histology and Anatomical Pathology are included. In contrast to other quality standards such as ISO 9001:2008 and GLP (Good Laboratory Practice), BS EN ISO/IEC 17025 is the only internationally recognised standard for testing laboratories that specifically demonstrates technical competence and the ability to generate technically valid results.

A detailed list of accredited tests with associated methods and Standard Operating Procedures is available on our website www.idexx.co.uk/ukas, and the scope for the accreditation can be found on the UKAS website www.ukas.org/testing.

Technically valid results are ensured via the use of:

• Validated testing methods, using suitable reagents, and equipment which is regularly calibrated, checked and maintained to a high standard

• Controlled Standard Operating Procedures (SOPs) for all test methods.

• Extensive quality control programmes including inter-laboratory comparisons.

Reference ranges provided with results for guidance are based on studies carried out in the laboratory wherever possible, but also take into account published information (which can reflect different methodologies and study populations). These are constantly under review by the Clinical Pathology team using experience from reporting cases and updated information from the veterinary field, and in our opinion offer the best aid in interpretation of reported results. As with all reference ranges some normal animals will lie outside these ranges.

The Quality Management System also requires that the business is managed in such a way as to ensure customer requirements are foremost. It is crucial that we understand our clients’ expectations – we do this via customer surveys, courtesy calls and taking on board all comments received. If you would like to lodge a complaint or let us know how we could improve our service then please contact the customer support teams (contact details on back page). We aim to resolve problems and, where appropriate, implement improved procedures as soon as possible.

We will inform you of the outcome, including follow up contact, to ensure ongoing satisfaction. As a result, we put resources into those areas of our service that you consider to be of primary importance.

Anne-Marie Porritt
NDV Quality Manager
Postal Addresses

IDEXX Laboratories
Wetherby
Grange House
Sandbeck Way, Wetherby
West Yorkshire, LS22 7DN

Sample Delivery
Wetherby
IDEXX Laboratories, PO Box 4
Wetherby, West Yorkshire, LS22 4YY

IDEXX Laboratories
London
84 New Wanstead
London, E11 2SY

Point-of-care
Analysers
IDEXX Laboratories Ltd
Riverside House
Riverside Walk, Windsor
Berkshire SL4 1NA

IDEXX Laboratories
Lea Green
Unit 12 Lea Green Business Park
Lea Green Ind Estate
Eurolink
St Helens, WA9 4TR