

Supplies Information

Animal Foundation (Non Interpreted)				
NSAID CLIP		■		
Chem 10 CLIP		■		
Chem 15 CLIP		■		
Chem 17 CLIP		■		
Lyte 4 CLIP		■		
Small Animal Core				
Basic		■ ■		■
Standard		■ ■		■
Extended		■ ■		■ ■
Small Animal Problem Specific				
Fits and Faints		■ ■		■
Polyuria/Polydipsia		■ ■		■ ■
Geriatric		■ ■		■
Canine Vomiting/Pancreatitis		■ ■		■
Feline Under the Weather		■ ■		■
Cardiac		■ ■		■ □
Anaemia		■ ■		■
Equine Profiles				
Basic Profile		■ ■		■
Comprehensive Profile		■ ■		■
Fitness Profile		■ ■		■
Rabbit / Other Small Mammal / Avian / Reptile Profiles				
Basic Rabbit Profile		■ ■		■
Comprehensive Rabbit Profile		■ ■		■
Rabbit Infectious Disease Screen		■		
Basic Small Mammal Profile		■ ■		■
Comprehensive Small Mammal Profile		■ ■		■
Avian Basic Profile			■ ■	
Comprehensive Avian Profile			■ ■	
Basic Reptile Profile			■ ■	
Comprehensive Reptile Profile			■ ■	
Haematology				
Comprehensive Haematology		■		■
Coagulation Profile		■ ■		■
Microbiology				
Urine			■ ■	■
Faeces			■	
Skin	Hair pluck / Skin scrape			
Histology				
Histology				■
Histology & Cytology on Fluids		■		■ ■
Histology & Cytology on Smear			■	■
Histology & Culture & Sensitivity			□	■ ■
Histology & Complete Skin examination	Hair pluck / Skin scrape			
				■

*Note a blood smear is not included in non-interpreted screens

Blood and other Body Fluids



■ Potassium EDTA Tube

Used for the analysis of full blood counts. The tube contains a chelating agent, which acts as an anticoagulant and preserves cellular morphology.

■ Separated EDTA plasma

Required for all Cardiopet® proBNP assays

- Centrifuge the EDTA sample.
- Place separated plasma into a plain tube.
- Clearly label with the code

BNPE CANINE / BNPE FELINE



■ Serum Gel S/1.1 Tube

Used for biochemistry and endocrinology, and allows natural clotting action of the blood. The tubes require centrifugation before dispatch (refer to protocol notes). Spun gel tubes prevent in-transit haemolysis. The tubes are not suitable for haematology.



■ Serum Z/1.3 Tube

Used for biochemistry and endocrinology, and allows natural clotting action of the blood. The tubes are not suitable for haematology.



■ Citrate Tube

Do not pool citrated plasma samples. Can only be used for coagulation assays, contains an anticoagulant which stabilises the coagulation factors. The volume of anticoagulant important as liquid dilutes blood. When submitting samples for coagulation, after mixing the blood in the citrate tube, please separate citrated plasma into the plain tube supplied and freeze for transit.



■ Li-Heparin Tube

The preferred sample tube for haematology analysis in exotic species. Contains Anti-Thrombin Anti-Xa-IIa which prevents coagulation. Good for exotic species as red cells are not lysed as with EDTA, however can lead to background staining in films.

Histology



■ Histology Pot (containing 10% NeutralBuffered formalin)

Specimens should be fixed in at least 10 times the volume of formalin relative to the size of the tissue. Small samples such as endoscopic biopsies are best submitted in mesh cassettes – these are available through our Supplies Department. Histology pots are provided in two sizes – small (40 ml pot containing 20 ml formalin) and large (120 ml pot containing 60 ml formalin). When submitting multiple biopsies, please use a separate pot for each tissue. For very large specimens, packaging supplies can be ordered online or through at idexx.co.uk/supplies.

Please observe the hazard information and implement the necessary arrangements for the safe use/handling use/handling, storage and~emergencies e.g. spillages.

Supplies Information

Microbiology



Plain Universal Container

Used for the collection of urine for microscopic examination for e.g. urinary sediment examination, urine chemistries and measurement of specific gravity. Also suitable for transport of cystocentesis samples for culture.



Boric Acid Container

Recommended for collection of urine for microbiological culture when not collected by cystocentesis. The presence of boric acid prevents bacteria overgrowth.



Faecal Container with Spoon

Required for the examination of faecal samples. If anaerobic examination is required the faecal container should be filled to the top to prevent too much air getting into the sample and reducing the number of viable anaerobic bacteria present. Too little sample may result in drying of the sample making recognition of any parasites present very difficult.



E-swab

Required for sample submission for microbiological examination and for PCR. Swabs should be collected from fresh, active lesions if possible. If deeper seated infection is likely, then the surface should be cleaned and disinfected, and when the site is dry, samples collected from deeper tissue.



Minitip e-swab

Replaces the charcoal minitip swab, which is popular for sampling those harder to reach areas and for smaller patients. Can be used for culture and PCR.



CEM Swab

Required for submission of swabs for contagious equine metritis (CEM), *Klebsiella pneumoniae* and *Pseudomonas aeruginosa*. Swab samples must arrive at the Wetherby laboratory within 48 hours of collection, and be accurately labelled with the horse's name, swab site, and the time and date of sampling.

Fresh tissue

Required for the submission of Microbiology in conjunction with Histology.

Others

Slides and Container



Fixed EDTA

Add a single drop of 10% neutral buffered formalin to an EDTA tube. Use for the submission of fluid aspirates. Aids the preservation of the cell morphology.

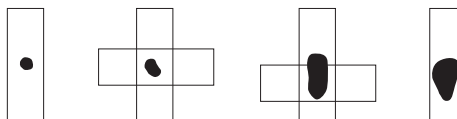
Sediment Smear

Required for the submission of smears for cytological analysis.

Preparation Guide

A small amount of the sample is placed on one slide and a second slide placed on top. The second slide may be perpendicular or parallel to the first. The material will spread between the two slides (Figure a). If necessary, very gentle pressure may be used to facilitate spreading (Figure b) and the top slide is gently pulled across the bottom until the two slides are separated (Figures c and d). The slides should slide apart, and should not be lifted away from each other. Rapid drying of the slides gives the best preservation of cellular morphology. This may be achieved by directing a hairdryer (cool / warm settings, not hot) onto the back of the slide from a distance of approximately 6 – 8 inches.

Fig. a Fig. b Fig. c Fig. d

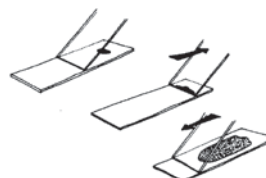


Wedge Smear

Required for the submission of smears for morphological examination

Preparation Guide

Place a small dot of whole (mixed) blood onto the slide. Place the spreader narrow side down onto the slide and pull back to the sample dot. The blood will spread across the width of the spreader. Push the spreader along the slide to form a smear. Do not lift the spreader off the slide until the entire sample has been spread.



To order supplies, please visit idexx.co.uk/supplies and complete the online order form.

If you would like any further information or assistance please contact a member of our Customer Support team:

Telephone: **UK:** +44 (0) 2037887508, **Eire:** +353 (0) 15621211 followed by Option 1

or email: labsupportUK@idexx.com